

Revised: February 23, 2022

Product Information

Phalloidin Conjugates

CF® Dye Phalloidin Conjugates

Catalog no. Unit Size Conjugate Ex/Em (nm) 00049-T 50 U Phalloidin, CF®350 347/448 00049-T 50 U Phalloidin, CF®405M 408/452 00034-T 50 U Phalloidin, CF®405M 408/452 00054-T 50 U Phalloidin, CF®430 426/498 00055-T 50 U Phalloidin, CF®430 440/515 00042-T 50 U Phalloidin, CF®480 490/515 00042-T 50 U Phalloidin, CF®488A 490/515 00051-T 50 U Phalloidin, CF®532 527/568 00051-T 50 U Phalloidin, CF®532 527/568 00043-T 50 U Phalloidin, CF®543 541/560 00044-T 50 U Phalloidin, CF®588 562/583 00044-T 50 U Phalloidin, CF®583R 586/609 00045-T 50 U Phalloidin, CF®683 593/614 00046-T 50 U Phalloidin, CF®660R 642/662 00050-T 50 U Phalloidin, CF®660R 642/662	or & Bye i nanolam conjugates				
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Other Phalloidin Conjugates

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Catalog no.	Unit Size	Conjugate	Ex/Em (nm)	
00028	100 U	Phalloidin, Biotin-XX	N/A	
00030	300 U	Phalloidin, Fluorescein	496/516	
00032	300 U	Phalloidin, Rhodamine 110	502/524	
00027	300 U	Phalloidin, Rhodamine	540/565	
00033	300 U	Phalloidin, Sulforhodamine 101 (Texas Red®)	591/608	

One unit of fluorescent phalloidin is defined as the amount of material used to stain one sample of fixed cells in a 200 uL volume (see protocols below).

Storage and Handling

Store at -20°C, desiccated, and protected from light. Lyophilized product is stable for at least one year from date of receipt when stored as recommended. After reconstitution in methanol or water, stock solutions are stable for at least one year when stored -20°C, protected from light. If using water as the solvent, freeze in aliquots. While the small amount of toxin in a vial is not likely to pose a heath hazard, it should be handled with care using universal laboratory safety precautions.

Important: See notes about the compatibility of specific CF® Dyes with fluorescence mounting media, and about the stability of phalloidin staining, after step 10 in the staining protocol (next page).

Product Description

Phalloidin is a toxin isolated from the deadly Amanita phalloides mushroom. It is a bicyclic peptide that binds specifically to F-actin (1). It is a very convenient tool to investigate the distribution of F-actin when labeled with fluorescent dyes. Phalloidin contains an unusual thioether bridge between cysteine and tryptophan residues that forms an inner ring structure. At elevated pH, this thioether is cleaved and the toxin loses its affinity for actin.

CF® Dyes are a series of next-generation fluorescent dyes developed at Biotium to have combined advantages in brightness, photostability, and water solubility compared to other fluorescent dyes. Fluorescently labeled phalloidins stain F-actin at nanomolar concentrations (1,3). Labeled phalloidins have similar affinity for both large and small filaments, binding in a stoichiometric ratio of about one phalloidin molecule per actin subunit in muscle and nonmuscle cells from various species of plants and animals. Different from antibodies, the binding affinity of phalloidin does not change significantly with actin among different species. Non-specific staining is negligible, and the contrast between stained and unstained areas is extremely large. Phalloidin shifts the monomer/polymer equilibrium toward the polymer, lowering the critical concentration for polymerization up to 30-fold (3,4). Phallotoxins also stabilize F-actin by inhibiting depolymerization caused by cytochalasins, potassium iodide, and elevated temperatures. Because the phalloidin conjugates are small, with an approximate diameter of 12-15 Å and molecular weight of <2000 Daltons, a variety of actin-binding proteins including myosin, tropomyosin and troponin can still bind to actin after treatment with phalloidin. Even more significantly, phalloidin-labeled actin filaments remain functional; labeled glycerinated muscle fibers still contract, and labeled actin filaments still move on solid-phase myosin substrates (5,6). Fluorescent phalloidin can also be used to quantify the amount of F-actin in cells (7,8).

Experimental Protocols

Preparation of Stock Solutions

CF® Dye phalloidin conjugates: dissolve the lyophilized solid in methanol or water (1.5 mL for the 300 U size or 0.25 mL for the 50 U size) to yield a stock solution of 200 U/mL.

Other fluorescent phalloidin conjugates: dissolve 300 U lyophilized solid in 1.5 mL methanol to yield a stock solution of 200 U/mL (approximately 6.6 uM).

Biotin-XX-phalloidin: dissolve 100 U lyophilized solid in 1 mL methanol to yield a stock solution of 100 U/mL (approximately 10 uM).

One unit (U) of fluorescent phalloidin is defined as the amount of material used to stain one microscope slide of fixed cells. For fluorescent phalloidins one unit is equivalent to 5 uL of 200 U/mL stock solution in a total staining volume of 200 uL. For biotin-XX-phalloidin, one unit is equivalent to 10 uL of 100 U/mL stock solution in a total staining volume of 200 uL.

Staining Fixed Cells

The following protocol describes the staining procedure for adherent cells grown on glass coverslips or 8-well chamber slides. Phalloidins also can be used to stain fixed frozen or paraffin tissue sections, as well as yeast and fungi.

Note: When staining yeast in liquid culture, cells in log phase stain much better than cells in stationary phase.

- 1. Wash cells 3 times with PBS.
- 2. Fix cells on ice with 3.75% formaldehyde solution in PBS for 15 minutes.

Note: Methanol can disrupt actin during the fixation process. Therefore, it is best to avoid any methanol containing fixatives or other solvent-based fixatives. The preferred fixative is methanol-free formaldehyde.

- 3. Wash 3 times with PBS.
- Permeabilize cells with 0.5% Triton X-100 in PBS at room temperature for 10 minutes.
- 5. Wash cells 3 times with PBS.
- Dilute 5 uL fluorescent phalloidin stock solution in 200 uL PBS for each cover slip or chamber to be stained. For biotin-XX-phalloidin, dilute 10 uL stock solution in 200 uL PBS. Volumes can be scaled as necessary depending on the size of the specimen or culture vessel.

Note: For staining yeast or fungi, increasing the phalloidin concentration from 5 U/mL to 50 U/mL may improve penetration into the cells.

Place the staining solution on the coverslip for 20 minutes at room temperature. To avoid evaporation, keep the coverslips inside a covered container and the chamber slides covered during the incubation.

Note: Phalloidin conjugates also can be included with fluorescently-labeled antibodies in blocking buffer during the secondary antibody incubation step in your regular immunofluorescence staining protocol.

- Wash 2-3 times with PBS.
- For biotin phalloidin, continue with biotin detection using labeled streptavidin or anti-biotin antibody. For fluorescent phalloidins, proceed to imaging.
- CF® Dye phalloidins can be imaged in PBS, but for best results, especially for preserving staining long-term we recommend mounting with EverBrite™ antifade mounting medium.

Note: CF®647, CF®660C, and CF®680 are cyanine-based dyes and are not compatible with VECTASHIELD® mounting media (Vector Labs.). Biotium's EverBrite™ antifade mounting media (see Related Products) are compatible with a wide-range of fluorescent dyes, including cyanine dyes and CF® Dyes.

Note: Fluorescent dyes can affect the stability of phalloidin staining which can cause loss of signal intensity over time. If samples are not mounted, it is highly recommended to image the cells immediately. For best results, store phalloidin-stained samples in a suitable mounting medium at 4°C, protected from light. For certain phalloidin conjugates, especially CF®543, CF®647, and CF®680, we recommend imaging immediately or shortly after staining. CF®647 and CF®680 phalloidins are recommended for STORM applications, but due to the instability of staining with these conjugates, we do not recommend using them for other microscopy applications. Staining with other CF® Dye phalloidin conjugates is more stable and signal could last for several days when specimens are stored at 4°C, protected from light.

Staining Living Cells

Fluorescently-labeled phalloidin is not cell-permeant and has therefore not been used extensively with living cells. However, living cells have been labeled by pinocytosis or unknown mechanisms (9-12). In general, a larger amount of stain will be needed for staining living cells. Alternatively, fluorescent phalloidins have also been injected into cells for monitoring actin distribution and cell motility (13-16).

References

1) Springer-Verlag New York. 47(1986); 2) J Muscle Res Cell Motil. 9, 370(1988); 3) Methods Enzymol. 85, 514(1982); 4) Eur J Biochem. 165, 125(1987); 5) Nature. 326, 805(1987); 6. Proc Natl Acad Sci USA. 83, 6272(1986); 7) Blood. 69, 945(1987); 8) Anal Biochem. 200, 199(1992); 9) J Cell Biol 105. 1473(1987); 10) Proc Natl Acad Sci USA. 77, 980(1980); 11) Nature. 284, 405(1980); 12) CRC Crit Rev Biochem. 5, 185(1978); 13) J Cell Biol. 106, 1229(1988); 14) J Cell Biol. 103, 265a(1986); 15) Eur J Cell Biol. 24, 176(1981); 16) Proc Natl Acad Sci USA. 74, 5613(1977).

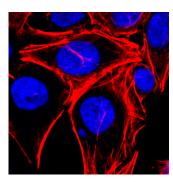


Figure 1. HeLa cells were fixed, permeabilized and stained with phalloidin, CF®640R conjugate (red) and DAPI (blue).

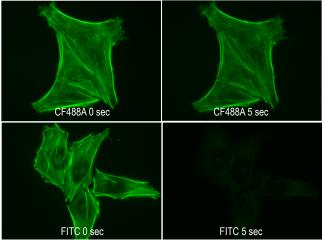


Figure 2. Relative photostability of CF®488A compared to FITC. HeLa cells were stained with CF®488A or FITC phalloidin conjugates and continuously exposed under a 100X objective using a mercury arc lamp microscope. Images were captured at t=0 and t=5 seconds of photobleaching.

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22006	Super ^{HT} Pap Pen 4 mm tip, ~800 uses		
22015	Fixation Buffer		
22016	Permeabilization Buffer		
22017	Permeabilization and Blocking Buffer		
22010	10% Fish Gelatin Blocking Buffer		
23007	TrueBlack® Lipofuscin Autofluorescence Quencher, 20X in DMF		
22014	Bovine Serum Albumin 30% Solution		
22002	Tween® 20		

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